

Antimicrobial Susceptibility of 27,123 Pathogens Isolated from Patients in Canadian Hospitals: CANWARD Study 2007 - 2011



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UPDATED ABSTRACT

Background: CANWARD is a national, annual, ongoing study assessing pathogens causing infections in Canadian hospitals and their antimicrobial resistance patterns.

Methods: From 2007 through 2011, 27,123 pathogens were collected from tertiary-care centres from across Canada. Susceptibility testing was performed using CLSI broth microdilution methods.

Results: Of the 27,123 isolates collected in total, 45.2%, 29.6%, 14.8% and 10.4% were from blood, respiratory, urine and wound specimens, respectively. Patient demographics were as follows: 54.4/45.6% male/female, 12.8% ≤17yrs, 45.1% 18-64yrs and 42.1% ≥65yrs. Isolates were obtained from patients in medical and surgical wards 37.8%, emergency rooms 25.7%, clinics 18.0% and ICUs 18.5%.

The most common pathogens were: E. coli 20.1%, S. aureus (MSSA) 15.4%, P. aeruginosa 8.0%, S. pneumoniae (SPN) 6.9%, K. pneumoniae 6.1%, Enterococcus spp. 5.9%, MRSA 4.7%, H. influenzae 3.8%, and Enterobacter cloacae 2.3%. Susceptibility rates (SR) for E. coli were: 100% meropenem (MER), 99.7% ertapenem (ERT), 97.7% piperacillin/tazobactam (P/T), 99.9% tigecycline (TGC), 93.7% ceftazidime (CTR), 90.5% gentamicin (Gent), 77.9% ciprofloxacin (CIP) and 73.4% cotrimoxazole (T/S). SR for P. aeruginosa were: 93.1% colistin, 82.6% MER, 84.0% P/T, 83.5% ceftazidime (CAZ), 72.0% Gent and 71.9% CIP. SR for MRSA were: 100% daptomycin (DAP), 100% linezolid (LZD), 100% telavancin (TLV), 99.8% TGC, 99.9% vancomycin, 92.2% T/S and 48.2% clindamycin. Statistical analysis revealed that ESBL rates among E. coli and K. pneumoniae and VRE rates increased while MRSA rates declined over time. Fluoroquinolone resistance rates in E. coli also increased over time.

Conclusions: E. coli, MSSA, P. aeruginosa, SPN, K. pneumoniae, MRSA and Enterococcus spp. are the most common pathogens in Canadian hospitals. SR for E. coli were highest with MER, ERT P/T and TGC. SR for P. aeruginosa were highest with colistin, MER, P/T and CAZ. 100% susceptibility occurred in MRSA to DAP, LZD and TLV with 99.9% with vancomycin.

INTRODUCTION

Pathogens causing antibiotic resistant infections is a Canadian and global crisis (1,2). Antibiotic resistant pathogens including methicillin-resistant Staphylococcus aureus (MRSA), community-associated and healthcare-associated), vancomycin-resistant Enterococcus species (VRE), penicillin-resistant Streptococcus pneumoniae (PRSP), extended spectrum β-lactamase (ESBL) producing Escherichia coli and Klebsiella species and fluoroquinolone-resistant and carbapenem-resistant Enterobacteriaceae and Pseudomonas aeruginosa are increasing in prevalence in Canada and around the world (1,2). Available therapeutic options for the treatment of these antibiotic resistant organisms are severely limited as these organisms frequently display a multidrug resistant (MDR) phenotype.

PURPOSE

- To determine the pathogens associated with respiratory, urinary, bacteremic and wound/IV site infections in Canadian patients affiliated with hospitals from 2007-2011, inclusive.
• To determine the prevalence of antimicrobial resistance in pathogens associated with respiratory, urinary, bacteremic and wound/IV site infections in Canadian patients affiliated with hospitals from 2007-2011, inclusive.
• To assess the activity of antimicrobials against respiratory, urinary, bacteremic and wound/IV site pathogens in Canadian patients affiliated with hospitals from 2007-2011, inclusive.

MATERIALS & METHODS

Participating Sites: From January 2007 to December 2011, sentinel hospital sites (12 in 2007, 10 in 2008, 15 in 2009, 14 in 2010 and 15 in 2011) in major population centres in 8 of the 10 provinces in Canada were recruited (1,2). These sites were geographically distributed in a population based fashion.

Bacterial Isolates: Tertiary-care medical centres submitted pathogens from patients attending hospital clinics, emergency rooms, medical and surgical wards, and intensive care units. From January 2007 through December 2011, inclusive, each study site was asked to submit clinical isolates (consecutive, one per patient, per infection site) from inpatients and outpatients with respiratory, urine, wound, and bloodstream infections. The medical centres submitted "clinically significant" isolates from patients with a presumed infectious disease. Surveillance swabs, eye, ear, nose and throat swabs were excluded. We also excluded anaerobic organisms. Isolate identification was performed by the submitting site and confirmed at the reference site as required, based on morphological characteristics and antimicrobial susceptibility patterns. Isolates were shipped on Amies semi-solid transport media to the coordinating laboratory (Health Sciences Centre, Winnipeg, Canada), subcultured onto appropriate media, and stocked in skim milk at -80°C until minimum inhibitory concentration (MIC) testing was carried out. In 2007, 2008, 2009, 2010 and 2011; 7714, 5283, 5372, 4960 and 3794 isolates were collected, respectively (1,2).

Antimicrobial Susceptibilities: Following 2 subcultures from frozen stock, the in vitro activity of selected antimicrobials was determined by broth microdilution in accordance with the Clinical and Laboratory Standards Institute (CLSI) guidelines (CLSI, 2009 M7-A8). Antimicrobial minimum inhibitory concentration (MIC) interpretive standards were defined according to CLSI breakpoints (M100-S21, 2011). Susceptibility testing could not be performed with all agents due to lack of space on the susceptibility panels. Antimicrobial agents were obtained as laboratory grade powders from their respective manufacturers. Stock solutions were prepared and dilutions made as described by CLSI (M7-A8, 2009). The MICs of the antimicrobial agents for the isolates were determined using 96-well custom designed microtitre plates. These plates contained doubling antimicrobial dilutions in 100µl/well of cation adjusted Mueller-Hinton broth and inoculated to achieve a final concentration of approximately 5 x 10^5 CFU/ml then incubated in ambient air for 24 hours prior to reading. Colony counts were performed periodically to confirm inocula. Quality control was performed using ATCC QC organisms including: S. pneumoniae 49619, S. aureus 29213, E. faecalis 29212, E. coli 25922, and P. aeruginosa 27853.

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Table 1. Antimicrobial activity against the most common Gram-positive cocci isolated from Canadian hospitals

Table with 5 columns: Organism (no. tested) / Antimicrobial Agent, % S, % I, % R, MIC (µg/mL) (MIC50, MIC90, Range). Rows include Staphylococcus aureus, Streptococcus agalactiae, Streptococcus pneumoniae, Streptococcus faecalis, and Enterococcus faecium.

Bacterial Isolates Collected
27,123 clinical isolates were collected for CANWARD 2007-2011.

- 12,261 (45.2%) were from blood, 8,020 (29.6%) from respiratory sources, 4,012 (14.8%) were from urine and 2,830 (10.4%) were from wounds
• 14,744 (54.4%) collected from male patients; 12,379 (45.6%) female patients

Table with 5 columns: Organism (no. tested) / Antimicrobial Agent, % S, % I, % R, MIC (µg/mL) (MIC50, MIC90, Range). Rows include Haemophilus influenzae, Klebsiella pneumoniae, Klebsiella oxytoca, Proteus mirabilis, and Stenotrophomonas maltophilia.

Table 2. Antimicrobial activity against the most common Gram-negative bacilli isolated from Canadian hospitals

Table with 5 columns: Organism (no. tested) / Antimicrobial Agent, % S, % I, % R, MIC (µg/mL) (MIC50, MIC90, Range). Rows include Acinetobacter baumannii, Enterobacter cloacae, Proteus mirabilis, Klebsiella pneumoniae, Klebsiella oxytoca, Proteus mirabilis, and Stenotrophomonas maltophilia.

Table 3. The 20 most common organisms isolated from Canadian hospitals

Table with 4 columns: Rank, Organism, n, % of Total. Lists top 20 organisms like E. coli, S. aureus, P. aeruginosa, etc.

RESULTS

Table with 5 columns: Organism (no. tested) / Antimicrobial Agent, % S, % I, % R, MIC (µg/mL) (MIC50, MIC90, Range). Rows include Haemophilus influenzae, Klebsiella pneumoniae, Klebsiella oxytoca, Proteus mirabilis, and Stenotrophomonas maltophilia.

For tigecycline, FDA approved breakpoints used: S, susceptible; I, intermediate; R, resistant; colistin (Polymyxin E); AMC, amoxicillin clavulanic acid; TZP, piperacillin/tazobactam; SXT, trimethoprim/sulfamethoxazole

CONCLUSIONS

- Of the 27,123 pathogens obtained, the most common were: E. coli 20.1%, S. aureus (MSSA) 15.4%, P. aeruginosa 8.0%, S. pneumoniae 6.9%, K. pneumoniae 6.1%, MRSA 4.7%, H. influenzae 3.8% and Enterococcus spp. 5.9%.
• Isolates were obtained from blood (45.2%), respiratory (29.6%), urine (14.8%) and wound specimens (10.4%), and from patients on medical wards 28.8%, surgical 9.0%, ER 25.7%, clinics 18.0% and ICU 18.5%.
• Susceptibility rates for E. coli were: 100% meropenem, 99.9% doripenem and tigecycline, 99.7% ertapenem, 99.6% amikacin, 97.7% piperacillin/tazobactam, 97.7% ceftazidime, 95.2% ceftazidime, 93.7% ceftazidime, 90.5% gentamicin, 77.9% ciprofloxacin and 73.4% trimethoprim/sulfamethoxazole.
• Susceptibility rates for P. aeruginosa were: 93.1% colistin, 90.1% amikacin, 84.2% doripenem, 84.0% piperacillin/tazobactam, 83.5% ceftazidime, 82.6% meropenem, 72.0% gentamicin and 71.9% ciprofloxacin.
• Susceptibility rates for MRSA were: 100% daptomycin, linezolid and telavancin, 99.9% vancomycin, 99.8% tigecycline, 92.2% trimethoprim/sulfamethoxazole, 48.2% clindamycin, 13.7% ciprofloxacin and 12.2% for clarithromycin.
• Statistical analysis revealed that ESBL rates among E. coli and K. pneumoniae and VRE rates increased while MRSA rates declined over time. Fluoroquinolone resistance rates in E. coli also increased over time.